



ISPKH26 PKH26 Red Fluorescent Cell Linker Kit

Product Description

The PKH26 Fluorescent Cell Linker Kits use proprietary membrane labeling technology to stably incorporate a yellow–orange fluorescent dye with long aliphatic tails (PKH26) into lipid regions of the cell membrane.^{1,2} The labeling vehicle provided in the kits (Diluent C) is an aqueous solution designed to maintain cell viability, while maximizing dye solubility and staining efficiency during the labeling step. Diluent C is iso–osmotic for mammalian cells and contains no detergents or organic solvents, but also lacks physiologic salts and buffers. The appearance of labeled cells may vary from bright and uniform to punctate or patchy, depending on the cell type being labeled and the extent to which membrane internalization occurs after labeling. However, PKH26 fluorescence is independent of pH within physiologic ranges and fluorescence intensity per cell is typically unaffected by the pattern of dye localization.

PKH26 fluoresces in the yellow–orange region of the spectrum (see Figure 1) and has been found to be useful for in vitro and in vivo cell tracking applications in a wide variety of systems. PKH26 is compatible with violet, green, red, or far–red viability probes used in cytotoxicity assays and with fluorescent proteins, antibodies, or DNA dyes emitting in these spectral regions. One of the most common uses of PKH26 is proliferation analysis based on dye dilution. Applications of dye dilution proliferation analysis include estimation of antigen–specific precursor frequencies and identification of quiescent or slowly dividing stem or progenitor cells in normal and neoplastic tissues.^{16–20} PKH26 has also proven useful for monitoring: uptake of exosomes, viruses, platelets, and other nanoparticles; apportionment of membrane in stem cell division; cell–cell membrane transfer; phagocytosis; antigen presentation; adhesion; signaling through gap junctions, and neuronal migration in tissue slices. Due to its extremely stable fluorescence, PKH26 is the cell linker dye of choice for in vivo cell tracking studies, particularly when labeled cells are to be followed for periods longer than a few weeks.

Components

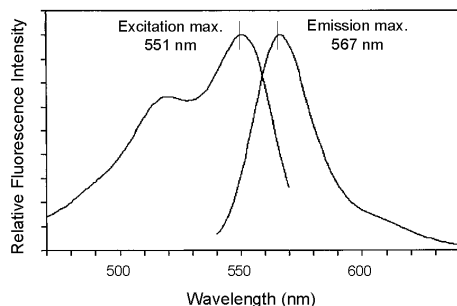
NO.	PKH26Dye	Diluent C
PKH26–0.1mL	0.1mL	10mL
PKH26–1mL	1mL	60mL
PKH26–5mL	5mL	120mL

Storage/Stability

The PKH26 ethanolic dye solution may be stored at room temperature or refrigerated. To prevent increases in dye concentration due to evaporation, keep the ethanolic dye solution tightly capped except when in immediate use. The dye solution must be protected from bright direct light and examined for crystals prior to use. If crystals are noted in the dye solution, warm slightly in a 37 °C water bath, and sonicate or vortex until



redissolved. Diluent C may be stored at room temperature or refrigerated. If refrigerated, bring to room temperature before preparing cell and dye suspensions for labeling. Diluent C is provided as a sterile solution. Because it does not contain any preservatives or antibiotics, it should be kept sterile. Do not store dye in Diluent C. Working solutions of dye in Diluent C should be made immediately prior to use.



Procedures

A. General Cell Membrane Labeling.

Labeling occurs by partitioning of the lipophilic dye into cell membranes. Labeling intensity is a function of both dye concentration and cell concentration and is not saturable. Therefore, it is essential that the amount of dye available for incorporation be limited. Over-labeling of cells will result in loss of membrane integrity and reduced cell recovery.

The following labeling procedure can be used for *in vitro* or *ex vivo* labeling of stem cells, lymphocytes, monocytes, endothelial cells, neurons, or any other cell type where partitioning of dye into lipid regions of the cell membrane is desired. Modified procedures may be required for *in vivo* labeling, for labeling platelets, or for selective labeling of phagocytes in the presence of non-phagocytes.

General cell membrane labeling should be performed prior to monoclonal antibody staining. The membrane dyes will remain stable during the monoclonal staining at 4 °C; however, capping of the monoclonal antibodies is highly probable if the general cell membrane labeling is carried out at ambient temperature subsequent to antibody labeling.

The cell and dye concentrations given in the following procedure represent starting concentrations that have been found broadly applicable to a variety of cell types. Users must determine the optimal dye and cell concentrations for their cell type(s) and experimental purposes by evaluating post-staining cell viability (e.g., propidium iodide exclusion), fluorescence intensity, staining homogeneity, and lack of effect on cell function(s) of interest.

Note: No azide or metabolic poisons should be present at the time of staining with PKH26.

Note: Although adherent cells may be labeled while attached to a substrate, more homogeneous staining is obtained using single cell suspensions. Best results will be obtained if adherent or bound cells are dispersed



into a single cell suspension using proteolytic enzymes, e.g., trypsin/EDTA, prior to staining.

The following procedure uses a 2 mL of final staining volume containing final concentrations of 2×10^{-6} M of PKH26 and 1×10^7 cells/mL.

Perform all further steps at ambient temperature (20–25 °C)

1. Place a suspension containing 2×10^7 single cells in a conical bottom polypropylene tube and wash once using medium without serum.

Note: Serum proteins and lipids also bind the dye, reducing the effective concentration available for membrane labeling. Best results are obtained by washing once with serum-free medium or buffer (step 1) prior to resuspension in Diluent C for labeling (step 4).

2. Centrifuge the cells (400 x g) for 5 minutes into a loose pellet.

Note: The PKH26 ethanolic dye solution should not be added directly to the cell pellet. This will result in heterogeneous staining and reduced cell viability.

3. After centrifuging cells, carefully aspirate the supernatant, being careful not to remove any cells but leaving no more than 25 μ L of supernatant.

Note: For reproducible results, it is important to minimize the amount of residual medium or buffer present when cells are resuspended in Diluent C.

4. Prepare a 2X Cell Suspension by adding 1 mL of Diluent C to the cell pellet and resuspend with gentle pipetting to ensure complete dispersion. Do not vortex and do not let cells stand in Diluent C for long periods of time.

Note: The presence of physiologic salts causes the dye to form micelles and substantially reduces staining efficiency. Therefore, it is important that the cells be suspended in Diluent C at the time dye is added, not in medium or buffered salt solutions.

5. Immediately prior to staining, prepare a 2x Dye Solution (4×10^{-6} M) in Diluent C by adding 4 μ L of the PKH26 ethanolic dye solution to 1 mL of Diluent C in a polypropylene centrifuge tube and mix well to disperse.

Note: To minimize ethanol effects on cell viability, the volume of dye added in step 5 should result in no more than 1~2% ethanol at the end of step 6.

Note: If a final dye concentration $< 2 \times 10^{-6}$ M is desired, the most reproducible results will be obtained by diluting the PKH26 ethanolic dye solution provided in the kit with undenatured 100% ethanol to make an intermediate dye stock.

6. Rapidly add the 1 mL of 2x Cell Suspension (step 4) to 1 mL of 2 x Dye Solution (step 5) and immediately mix the sample by pipetting. Final concentrations after mixing the indicated volumes will be 1×10^7 cells/mL and 2×10^{-6} M PKH26.



Note: Because staining is nearly instantaneous, rapid and homogeneous dispersion of cells in dye solution is essential for bright, uniform, and reproducible labeling. The following measures have been found to aid in optimizing results:

- a. Do not add ethanolic PKH26 dye directly to the 2x Cell Suspension in Diluent C.*
- b. Mix equal volumes of 2x Cell Suspension (step 4) and 2 x Dye Solution (step 5).*
- c. Adjust 2 x cell and 2 x dye concentrations to avoid staining in very small (<100 μ L) or very large (>5 mL) volumes.*
- d. Use a Pipetman or equivalent for rapid addition of cells and mixing with dye. Serological pipettes are slower and give less uniform staining. Mixing by “racking” or vortexing is also slower and gives less uniform staining.*
- e. Dispense volumes as precisely as possible in order to accurately reproduce both cell and dye concentrations from sample to sample and study to study.*

7. Incubate the cell/dye suspension from step 6 for 1~5 minutes with periodic mixing. Because staining is so rapid, longer times provide no advantage.

Note: Expose cells to dye solution and Diluent C for the minimum time needed to achieve the desired staining intensity. Since Diluent C lacks physiologic salts, longer exposure may cause reduced viability in some cell types. If such effects are suspected, include a diluent-only control and a mock-stained control using ethanol rather than dye.

8. Stop the staining by adding an equal volume (2 mL) of serum or other suitable protein solution (e.g., 1% BSA) and incubate for 1 minute to allow binding of excess dye.

Note: Serum (or an equivalent protein concentration) is preferred as the stop solution. Increase volume to 10 mL if complete medium is used instead of serum.

Note: Do not stop by adding Diluent C or centrifuge the cells in Diluent C before stopping the staining reaction.

Note: Do not use serum-free medium or buffered salt solutions, which cause formation of cell-associated dye aggregates. Dye aggregates act as slow-release reservoirs of unbound dye that are not efficiently removed by washing and can transfer to unlabeled cells present in an assay.

9. Centrifuge the cells at 400 x g for 10 minutes at 20~25 °C and carefully remove the supernatant, being sure not to remove cells. Resuspend cell pellet in 10 mL of complete medium, transfer to a fresh sterile conical poly-propylene tube, centrifuge at 400x g for 5 minutes at 20~25 °C, and wash the cell pellet 2 more times with 10 mL of complete medium to ensure removal of unbound dye.

Note: Transfer to a fresh tube increases washing efficiency by minimizing carryover of residual dye bound to tube walls.

Note: Do not use Diluent C for washing steps.



10. After the final wash, resuspend the cell pellet in 10 mL of complete medium for assessment of cell recovery, cell viability, and fluorescence intensity. Centrifuge and resuspend to desired final concentration of viable cells.
Note: Stained cells may be fixed with 1–2% neutral buffered formaldehyde and intensities are stable for at least 3 weeks if samples are protected from light.

Note: Staining is typically at least 100–1,000 times brighter than background autofluorescence. Intensity distributions should be symmetrical and as homogeneous as possible, although staining CV will depend on the cell type being stained.

B. Histology

Using DAB photo-oxidation and transmission electron microscopy it is possible to visualize internalization of PKH26 labeled membranes or nanoparticles at the ultrastructural level. PKH26 labeled adipocytes have been successfully identified in tissues subjected to standard paraffin embedding and sectioning, but such methods risk loss of intensity because they use clearing agents that may partially extract membrane lipids and lipophilic dyes.

Histologic studies of tissues containing cells labeled with lipophilic membrane dyes have typically been carried out on serial frozen sections or sections prepared after fixation in neutral buffered formalin prepared from 4% paraformaldehyde. These methods avoid quenching of fluorescence by absorbing dyes found in histologic counterstains. For studies where imaging is to be done on a single section, fluorescence microscopy should precede counterstaining.

The following methods were developed at the Pittsburgh Cancer Institute by Drs. Per Basse and Ronald H. Goldfarb for use with PKH26.

Fluorescence imaging of frozen sections:

1. Excise tissues to be sectioned and freeze immediately on dry ice.
2. Store tissues at -70°C prior to sectioning.
3. Mount frozen tissues using O.C.T. compound (Tissue-Tek; Miles, Inc.) or equivalent.
4. Prepare tissue sections.
5. Air dry slides for at least 1 hour at room temperature.
6. Mount coverslip using 1–2 drops of cyanoacrylate ester glue.
7. Examine or photograph sections using an appropriate filter setup (e.g., PE for PKH26).

Counterstaining of frozen sections:

1. Remove coverslips by soaking slides in acetone for 24–48 hours.
2. Rinse slides in distilled water to remove acetone.



3. Counterstain sections using stain of choice. Satisfactory results have been obtained using Mayer's or Harris hematoxylin.
4. Mount slides using AS/AP permanent aqueous mounting medium (Bio/Can America, Inc., Portland, ME) or equivalent.

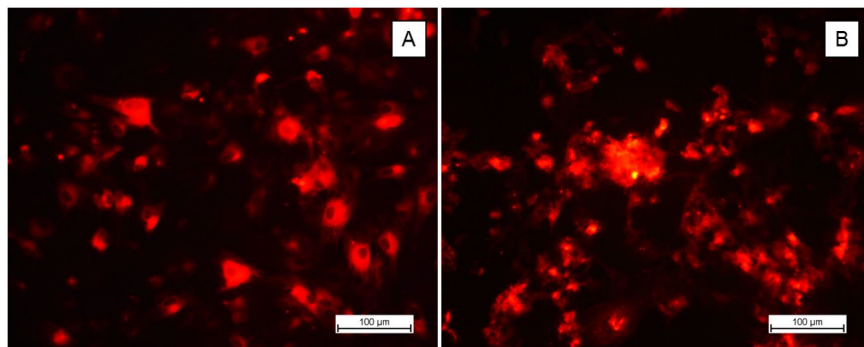
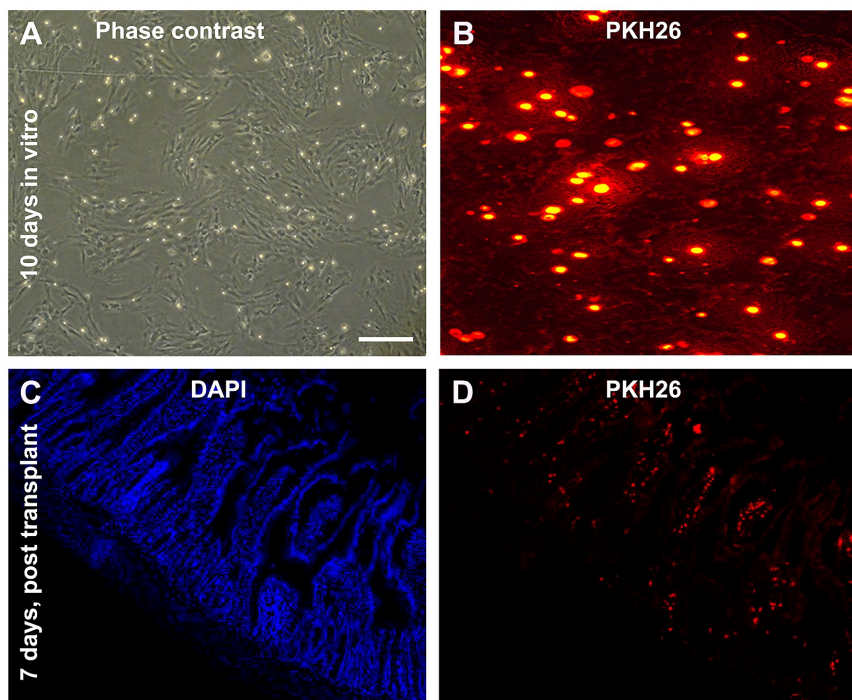
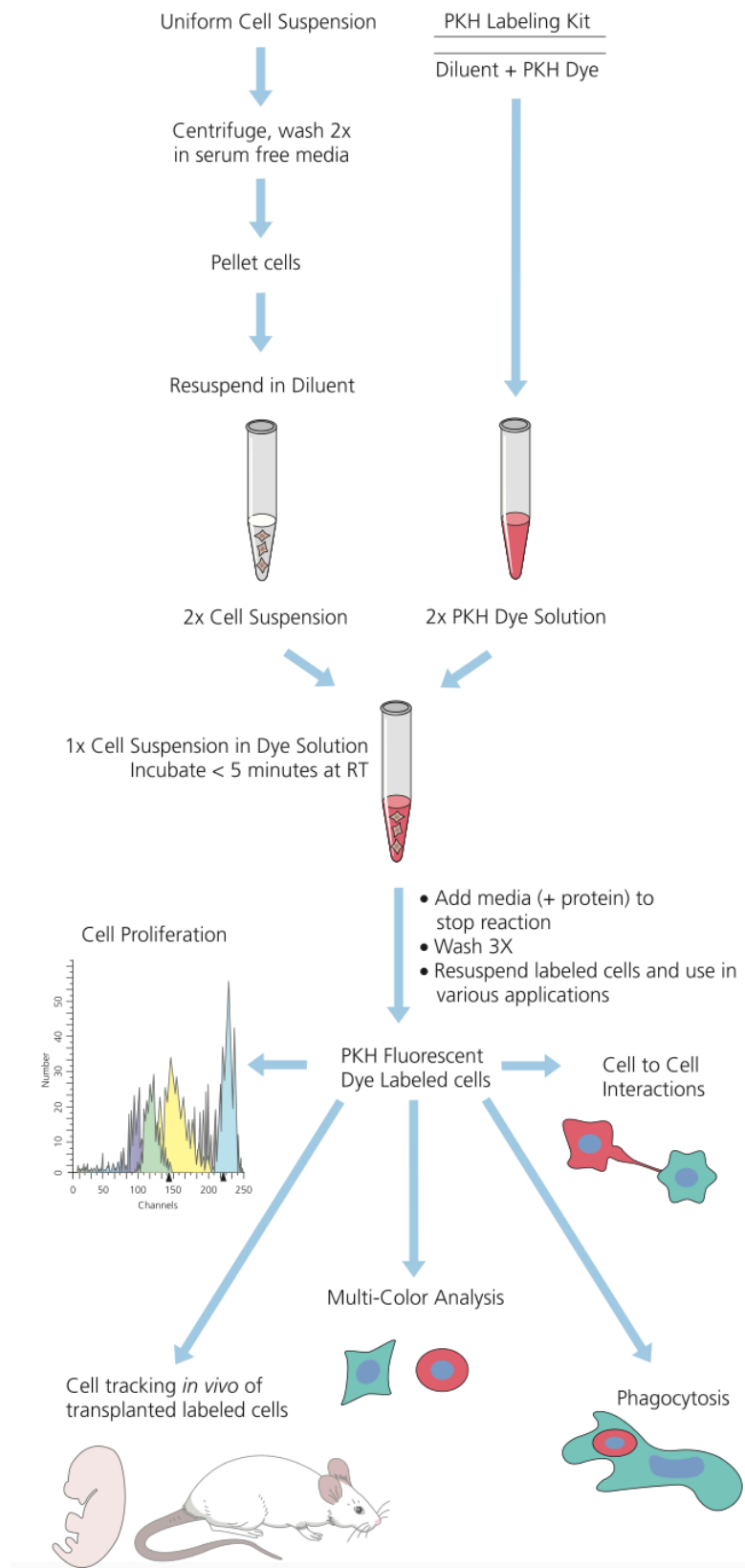


Figure Confirmation of cell labelling with the fluorescent dye, PKH26. (A) Osteoblastic cells grown on glass coverslips after 24 h; (B) osteoblast cell culture after 14 days



(A) A large number of cells adhering to a culture dish with spindle–shape morphology at 10 days in vitro. (B) Culture cells pre–labeled by PKH26 show bright red fluorescence. Panels (C) and (D) show the distribution of PKH26 positive BMMSCs in the wall of the small intestine 7 days after infusion, with the cells mostly localized to the mucosal region. Scale bar =500 µm in (A) applying for (C, D), equivalent to 125 µm for (B).



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