

# CO-F018 URIC ACID Fluorimetric Assay kit

Fluorescence microplate reader: (Ex/Em=535 nm/587 nm)

Size: 96T (Can delect 80 samples without duplication)

Sensitivity: 0.03 µmol/L

Detection range: 0.03-15 µmol/L

Average intra-assay CV(%): 1.5

Average inter-assay CV(%): 7.2

Average recovery rate(%): 101

#### - Intended use

This kit can be used to measure the Uric Acid (UA) content in urine, serum, plasma and animal tissue samples .

#### - Background

Uric acid, a purine metabolite, is degraded into allantoin by uricase in most mammals. Due to the absence of urate oxidase gene, uric acid is the final product of purine metabolism in humans, so the level of urie acid in human blood is higher than that in most mammals. Urie acid is a physiologically important plasma antioxidant that effectively protects biological targets from the oxidation of hydroxyl radicals, hypochloric acid and peroxynitrite.

# - Detection principle

Uricase catalyzes the decomposition of uric acid into allantoin, CO, and  $H_2O_2$  Under the action of peroxidase, H2O2 oxidizes the non-fluorescent probe into the fluorescent substance. By measuring the fluoresce value of the system , the corresponding uric acid content can be calculated.

#### Kit components & storage

| Item      | Component                       | Size             | Storage                          |  |
|-----------|---------------------------------|------------------|----------------------------------|--|
| Reagent 1 | Buffer Solution                 | 60 ml x 2 vials  | -20°C , 12 months                |  |
| Reagent 2 | Probe Solution                  | 0.24 ml x 1 vial | -20°C , 12 months, shading light |  |
| Reagent 3 | Enzyme Reagent 1                | 0.24 ml x 1 vial | -20°C , 12 months                |  |
|           | Enzyme Reagent 2                | 1.2 mlx 1 vial   | -20°C , 12 months                |  |
| Reagent 5 | 20 µmol/I Uric Acid<br>Standard | 1.5 mlx 1 vial   | -20°C , 12 months                |  |
|           | Black Microplate                | 96 wells         | No requirement                   |  |
|           | Plate Sealer                    | 2 pieces         |                                  |  |

Note: The reagents must be stored strictly according to the preservation conditions in the above table. The reagents in different kits cannot be mixed with each other.

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#### ·Safety data

Some of the reagents in the kit contain dangerous substances. It should be avoided to touch the skin and clothing. Wash immediately with plenty of water if touching it carelessly. All the samples and waste material should be treated according to the relevant rules of laboratory's biosafety.

#### - Precautions

Before the experiment, please read the instructions carefully, and wear gloves and work clothes.

#### Reagent preparation

- 1. Bring all the reagents lo room temperature belare use.
- 2. Preparation ol working solution:

Mix the reagent 1, reagent 2, reagent 3 and reagent 4 at the ratio of 36:2:2:10 lully. Prepare the Iresh solution belare use and stare with shading light.

#### -Sample preparation

The samples should be prepared as conventional methods. Also please refer to appendix II.

# **Dilution of sample**

Il is recommended lo lake 2-3 samples wilh expected large difference lo do pre-experiment before formal experiment and dilule the sample according to the result of the pre-experiment and the detection range (0.03-15 µmol/L).

The recommended dilution factor for different samples is as follows (for reference only):

| Sample type                      | Dilution Factor |  |  |
|----------------------------------|-----------------|--|--|
| Human serum                      | 10-20           |  |  |
| Human urine                      | 80-100          |  |  |
| Human hydrolhorax                | 50-60           |  |  |
| Rat urine                        | 10-20           |  |  |
| Rabbit serum                     | 5-10            |  |  |
| Rat serum                        | 10-20           |  |  |
| Porcine serum                    | 1               |  |  |
| 10% Rat liver tissue homogenate  | 10-20           |  |  |
| 10% Rat kidney tissue homogenate | 30-40           |  |  |
| 10% Rat lung tissue homogenate   | 10-20           |  |  |

Note: The diluent is reagent 1.

# **Assay Protocol**

# Plate Set up

|   | 1 | 2 | 3  | 4   | 5   | 6   | 7   | 8   | 9   | 10  | 11  | 12  |
|---|---|---|----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Α | Α | Α | S1 | S9  | S17 | S25 | S33 | S41 | S49 | S57 | S65 | S73 |
| В | В | В | S2 | S10 | S18 | S26 | S34 | S42 | S50 | S58 | S66 | S74 |
| С | С | С | S3 | S11 | S19 | S27 | S35 | S43 | S51 | S59 | S67 | S75 |
| D | D | D | S4 | S12 | S20 | S28 | S36 | S44 | S52 | S60 | S68 | S76 |
| Е | Ε | Е | S5 | S13 | S21 | S29 | S37 | S45 | S53 | S61 | S69 | S77 |
| F | F | F | S6 | S14 | S22 | S30 | S38 | S46 | S54 | S62 | S70 | S78 |
| G | G | G | S7 | S15 | S23 | S31 | S39 | S47 | S55 | S63 | S71 | S79 |
| Н | Η | Н | S8 | S16 | S24 | S32 | S40 | S48 | S56 | S64 | S72 | S80 |

Note: A-H, standard wells; S1-S80, sample wells.

# 'Detailed operation steps

The preparation of standard curve

Dilute 20  $\mu$ mol/L uric acid standard with reagent 1 to a serial concentration. The recommended dilution gradient is as follows: 15, 12,10, 8, 6, 4, 2, O  $\mu$ mol/L.

## Reference is as follows:

| N° | Standard concentration (µmol/L) | 10 (µmol/L)<br>Standard (µl) | Reagent (µI) |
|----|---------------------------------|------------------------------|--------------|
|    | 0                               | 0                            | 000          |
| Α  | U                               | U                            | 200          |
| В  | 2                               | 20                           | 180          |
| С  | 4                               | 40                           | 160          |
| D  | 6                               | 60                           | 140          |
| Е  | 8                               | 80                           | 120          |
| F  | 10                              | 100                          | 100          |
| G  | 12                              | 120                          | 80           |
| Н  | 15                              | 150                          | 50           |

The measurement of samples

- 1)Standard well: add 50 µL ol standard with different concentrations into the wells of 96 well microplate.
- 2) Sample well: add 50  $\mu$ L ol sample into the wells of 96 well microplate.
- 3) Add 50 µL of working solution into each well.
- 4) Mix fully with microplate reader lor 5 s and incubate al 37°C lor 30 min.
- 5) Measure the fluorescence intensity at the excitation wavelength of 587 nm.

# Summary operation table

|  | Standard<br>well | Sample<br>well |  |  |  |  |
|--|------------------|----------------|--|--|--|--|
| Standard with different concentrations (µL)  | 50               |                |  |  |  |  |
| Sample (µL)  |                  | 50             |  |  |  |  |
| Working solution (μL)  | 50               | 50             |  |  |  |  |
| Mix fully and and incubate at 37°C for 30 min. Measure the fluorescence intensity. |                  |                |  |  |  |  |

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## Calculation

Plot the standard curve by using fluorescence value (F) of standard and correspondent concentration as y-axis and x-axis respectively. Create the standard curve with graph software (or EXCEL). The concentration of the sample can be calculated according to the formula based on the F value of sample. The standard curve is: y= ax + b.

# Serum (plasma) and other liquid sample:

UA content  $(\mu \text{mol/L}) = (\Delta F - b) \div a \times f$ 

#### Tissue sample:

UA content ( $\mu$ mol/ gprot) = ( $\Delta$ F - b)  $\div$  a x f + C<sub>PR</sub>

#### Note:

- y: The absolute fluorescence value of standard, F<sub>stanadard</sub>- F<sub>blank</sub>, (F<sub>blank</sub> is the F value when the standard concentration is O)
- x: The concentration of standard.
- a: The slope of standard curve.
- b: The intercept of standard curve.
- ΔF: Absolute fluorescence intensity of sample (F<sub>sample</sub> F<sub>blank</sub>)
- f: The dilution factor of tested samples.
- C<sub>pr</sub>:The concentration of protein in sample, gprot/L

# **Example analysis**

Dilute 50  $\mu$ L of human urine with reagent 1 tor 100 times, take 50  $\mu$ L of diluted sample and carry the assay according to the operation table. The results are as follows:

standard curve: y = 227.73 x + 141.88, the average fluorescence value of the sample is 3077.9, the average fluorescence value of the blank is 277.3, and the calculation result is:

UA content ( $\mu$ mol/L) = (3077.9-277.3-141·88) = x 100=1167.49  $\mu$ mol/L

227.73

## Appendix II - Sample Preparation

#### Serum

Collect fresh blood and stand at 25°C lor 30 min lo clot the blood. Then centrifuge at 2000 g far 15 min at 4°C. Take the serum (which is the upper light yellow clarified liquid layer) to preserve it on ice far detection.

#### **Plasma**

Take fresh blood into the tube which has anticoagulant, centrifuge at 700-1000 g lor 10 min al 4  $^{\circ}$ C. Take the plasma (which is the upper light yellow clarified liquid layer, don'! take white blood cells and platelets in the middle layer) to preserve it on ice far detection.

#### Urine

Collect fresh urine and centrifuge at 10000 g lor 15 min at  $4^{\circ}$ C. Take the supernatant far delection. The UA has a low solubility and is easy to farm crystallization precipitation, so it should be heated to  $50^{\circ}$ C and then carry the assay.

### Tissue sample

Accurately weigh the tissue sample, add reagent 1 according to the ratio of Weight (g): Volume (ml) =1 :9. Mechanical homogenate the sample in ice water bath. Centrifuge at 10000 g far 10 min, then take the supernatant and preserve it on ice far detection. Meanwhile, determine the protein concentration of supernatant with BCA kit.

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# NOTE:

- 1. Homogenized medium: Reagent 1.
- 2.Homogenized method:
- (1)Hand-operated: Weigh the tissue and mince to small pieces (1 mm'), then put the tissues pieces to glass homogenized tube. Add homogenized medium into homogenized tube, piace the tube into the ice bath with left hand, and insert the glass tamping rod vertically into the homogenized tube with the right hand to grind up and down lor 6-8 min.

Or put the tissue into the mortar, and add liquid nitrogen to grind fully. Then add the homogenized medium to homogenize.

(2)Mechanical homogenate: Weigh the tissue to EP tube, add the homogenized medium to homogenize the tissue with homogenizer instrument (60 Hz, 90s) in the ice bath. (For samples of skin, muscle and plani tissue, the time of homogenization can be properly prolonged.)

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