Precast Gels Plus (Tris-Glycine)

compatible with Biorad Miniprotean Electrophoresis Tank

Introduction:

Our Plastic Precast Gels Plus (Tris-Gly) are precast polyacrylamide gels with high-performance and better band resolution.

- Automated gel casting technology can ensure stability and repeatability.
- Coated plastic plate can effectively reduce the non-specific adsorption of protein and make the protein band sharper and clearer.
- Gel cassettes can be easily opened with the Gel Opener.
- Precast Gels Plus do not contain SDS. Compatible with denaturing and native electrophoresis.
- Compatible with mainstream mini electrophoresis tanks on the market, such as Bio-Rad,
- Precast Gels Plus are available in different acrylamide concentrations, including gradient 4-15%、4-20% of separating gels.

Product Details：

- Height of 4% stacking gel: 1.5cm
- Ratio of acrylamide to methylene bis acrylamide: 29:1
- Gel thickness: 1.0mm
- Well format: 10wells/15wells
- Max sample loading volume: 30μL/well for 15wells, 50μL/well for 10wells
- Cassette size: 100×89×4.8 mm
- Gel size: 84×74×1 mm
- Storage: stored at 4℃ for 12 months; do not freeze, room temperature: 3 months

POLYACRYLAMIDE PRECAST GELS (TRIS GLYCINE) Suitable for Biorad Electrophoresis Machines.

CAT. #	GRADIENT	Wells	Max volume	Running buffer	Separation Range	Voltage
PSG2001-415	4-15%	10	50μL	Tris-Glycine	20-200kDa	180V
PSG2001-415	4-15%	15	30μL	Tris-Glycine	20-200kDa	180V
PSG2001-420	4-20%	10	50μL	Tris-Glycine	5-200kDa	180V
PSG2001-420	4-20%	15	30μL	Tris-Glycine	5-200kDa	180V

PRE CAST Gel Protocol：

Precast Gels Plus (Tris-Glycine) do not contain SDS and can be used for denaturing and non-denaturing electrophoresis depending on the running buffer.

A.For denaturing electrophoresis:

1. Prepare sample: Mix protein sample loading buffer for denaturing electrophoresis with sample in 1:4 ratio (volume) and then heat the samples at 95℃ for 5-10 minutes for optimal results.
2. Prepare running buffer for denaturing electrophoresis: Use **Tris-Glycine Running Buffer** for **SDS-PAGE** if you have 10× running buffer must be diluted to 1× final concentration before use. .

Note: The buffer system of Precast Gels Plus (Tris-Gly) is neutral buffer system.

3. Take the Precast Gels Plus (Tris-Gly) out of the bag and **remove the sealing tape from the top to the bottom of gel cassette**. Assemble the corresponding electrophoresis tank, add the running buffer, and then gently pull the comb out of the cassette.
4. Rinse the wells several times with 1 ×running buffer to remove storage buffer before loadingsamples. Load the appropriate volume of your samples in the appropriate wells. Load your protein ladder in the appropriate well.

Note: Do not insert the pipette tip too much into the sample wells to avoid sample leakage caused by deformation of the plastic plates.

5. Run the gel: The electrophoresis conditions are usually 180V, 50~60 min.

Note: The voltage is set at 180V constant with an initial current of 75 mA per gel, and a run time of about 60 minutes, and the current will gradually decrease with time.

6. After electrophoresis is complete, retrieve the gel from gel cassette: Inserting the Gel Opener between the two plates and push down gently on the knife handle to separate the plates; repeat on each side of the cassette until the plates are completely separated; then remove the top plate and gently peel away the gel from the plate.

Note: If the isoelectric point of the protein samples is greater than 7, you will need to reverse the electrodes for non-denaturing electrophoresis

Compatible electrophoresis tanks:

Precast Gels Plus (Tris-Gly) is compatible with most common mini SDS-PAGE tanks, including:

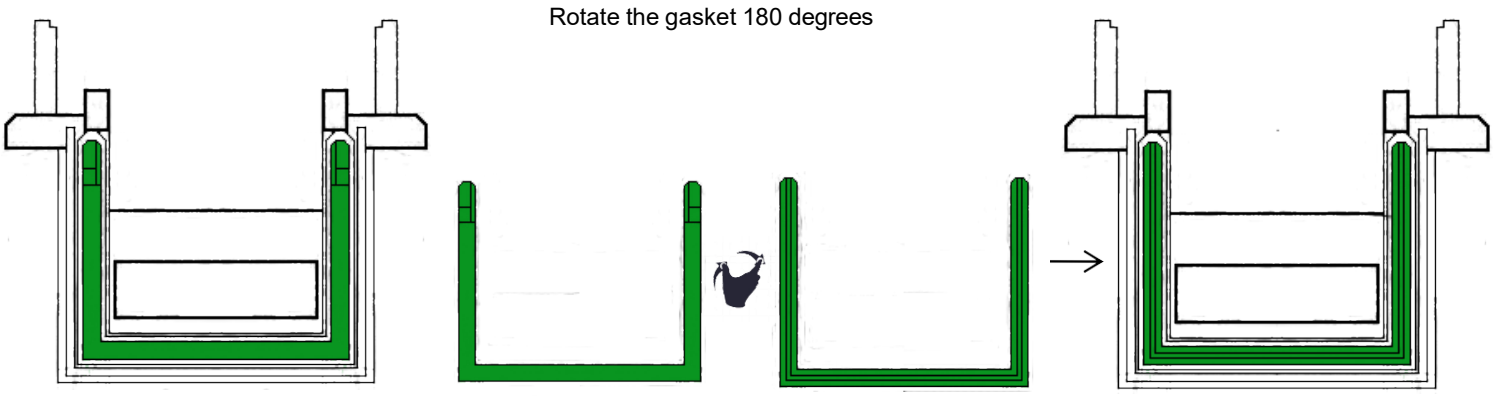
- **Bio-Rad Mini-PROTEAN (**a simple modification is required, see below)

Precast Gels Plus (Tris-Gly) used in Bio-Rad electrophoresis tank:

a. Pull out the U-shaped silicone gasket (green), electrode inner core silicone.

b. Rotate the gasket 180° degrees and install it back oppositely (outside inwards, inside-out). Press down around with your thumbs using even pressure to ensure it seal well to prevent leakage.

c. Now the system is ready to use.

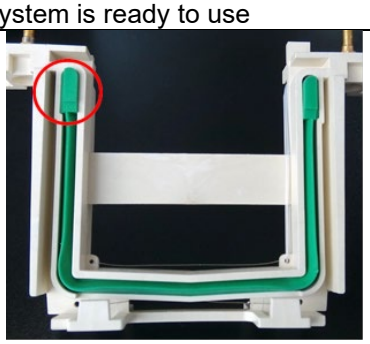


a. Pull out the U-shaped silicone gasket (green), electrode inner core Silicone (picture A.)

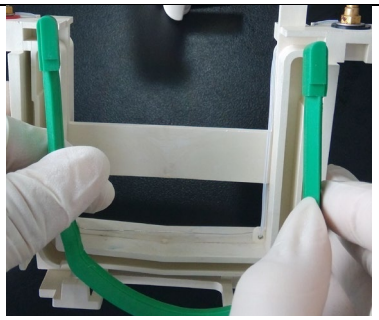
b. Rotate the gasket 180 degree and Installed it back oppositely (outside inwards, inside-out) picture B.

c. Press down around with your thumbs using even pressure to ensure to seal well to prevent leakage.

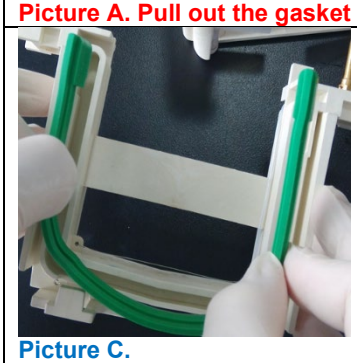
d. Now the system is ready to use



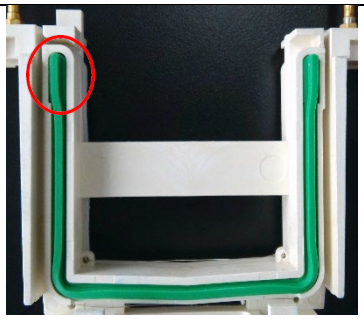
Picture A. Pull out the gasket



Picture B. Rotate 180°



Picture C.
Install it back opposite and press well to prevent any leakage



Picture D.
The system is ready

Distributed by:

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